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4 **Molecular Inclusion -Novel Approach to Enhance Solubility of Olmesartan Medoxomil using**  
5 **Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ -CD)**

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14 **ABSTRACT**

15 Olmesartan medoxomil (OLM) is a BCS Class II anti-hypertensive drug, which is having low aqueous  
16 solubility and low bioavailability of 26%. In the present study, the objective of the study was to  
17 improve the pharmaceutical limitation of OLM by using Molecular Inclusion Technique. Inclusion  
18 complex of OLM with Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ -CD) were prepared in drug/carrier molar  
19 ratios of 1:1, 1:2, and 1:3 by Solvent evaporation and Kneading method along with the Physical  
20 mixtures. The formation of molecular inclusion complex at 1:2 ratio in the solution was confirmed by  
21 Job's Plot study. The inclusion complex were characterized by Phase solubility study, Fourier  
22 transform infrared spectroscopy, Scanning electron microscopy, Differential scanning calorimetry and  
23 X-ray diffraction studies. *In- vivo* pharmacokinetic study reveals enhanced bioavailability of inclusion

24 complexes. Thus, it can be concluded that the formulation of molecular inclusion complex can  
25 enhance Solubility and Dissolution making the drug more suitable for treatment.

26 **Keywords:** Olmesartan medoxomil, Molecular Inclusion, Stability Constant, Dissolution, HP  $\beta$ -CD.

27

## 28 INTRODUCTION

29 In recent trends, there are many of newly synthesized active agents that suffer from low aqueous  
30 solubility and dissolution and therefore have low bioavailability. There are two approaches that focus  
31 on improving the oral bioavailability that is enhancement of solubility and dissolution rate and  
32 increasing permeability of such drugs. Solubilisation, salt formation, complexation, solid dispersion  
33 and size reduction are commonly used approaches to increase dissolution rate. Amongst all these,  
34 solid dispersion technology is the promising approach for improvement of solubility and dissolution  
35 rate of poorly water soluble drugs 1. Recently, Solid dispersion technology was explored extensively  
36 for the delivery of the drugs having poor solubility. Solid dispersions are mixtures in which drugs  
37 exist either in an amorphous form dispersed in the carrier molecule or included into the cavity of the  
38 carrier molecule 6. Molecular inclusion technology has been proved a promising form of drug  
39 delivery. It offers dispersion or inclusion of a hydrophobic drug in a hydrophilic matrix of a carrier  
40 molecule in order to improve the dissolution rate which eventually improves the bioavailability of the  
41 drug.

42 Cyclodextrin (CD) inclusion complexation has been proved as a promising technique in enhancing  
43 solubility of insoluble drugs. There is the formation of host-guest inclusion complexes by weak  
44 intermolecular interaction <sup>12</sup>. The structure of cyclodextrin appears as a truncated cone, which has  
45 hydrophilic exterior surface and a lipophilic interior cavity into which drug molecules get entrapped 8.  
46 Hydroxypropyl- $\beta$ CD (HP $\beta$ CD) is the commonly used excipients in several pharmaceutical  
47 formulations<sup>11</sup>. From the literature survey it was revealed that the solubility of  $\beta$ CD in water is  
48 relatively low (approximately 18.5 mg/mL at 25 °C), as compared to its derivative HP $\beta$ CD which has  
49 a higher aqueous solubility (approx. 600 mg/mL at 25 °C). The reason behind the higher solubility of

50 HP $\beta$ CD is the chemical modification in the  $\beta$ CD by the addition of hydroxy propylene oxide group. 4.  
51 Also, HP- $\beta$  cyclodextrin is most useful drug complexing agent because of its ability to readily form  
52 complexes and reasonable cost. Therefore, it was thought worthwhile to select HP $\beta$ CD as a carrier for  
53 the formation of molecular inclusion complex.

54 Olmesartan medoxomil is a specific angiotensin II type I antagonist used alone or with other anti-  
55 hypertensive agents to treat hypertension. Olmesartan has poor aqueous solubility and low  
56 bioavailability of 26% 9. In the recent study, it was also explored that OLM has a better renal  
57 protective effect than other Angiotensin receptor blockers. Therefore, improvement of solubility of  
58 OLM can prove this drug as a better option for the treatment of Hypertension for the patients suffering  
59 from renal disorder <sup>12</sup>. HP- $\beta$ CD forms inclusion complexes in both solid and solution state, which can  
60 lead to modification in physical as well as chemical properties of the guest molecule. Solid drugs can  
61 be complexed with HP- $\beta$ CD by using solvent evaporation, freeze drying, kneading, roll mixing etc.  
62 techniques.

63 The objective of the present study is to develop OLM-molecular inclusion complex with improved  
64 solubility using Hydroxypropyl- $\beta$  cyclodextrin (HP $\beta$ CD) as a carrier with various techniques like  
65 physical mixing, solvent evaporation and kneading. The study includes phase solubility study,  
66 characterization of inclusion complex by DSC and, SEM, determination of stability constant of the  
67 complex, solubilisation and stability studies of the product.

## 68 **METHODOLOGY**

### 69 **Materials**

70 Olmesartan medoxomil (Mylan Laboratories, Hyderabad), HP- $\beta$  Cyclodextrin (Kleptose<sup>®</sup> Roquette,  
71 France) and other reagents used were of AR Grade.

### 72 **Preparation of Inclusion Complexes**

73 1) Physical mixtures: The physical mixtures of Drug OLM and HP- $\beta$  Cyclodextrin in different ratios  
74 were prepared by mixing the pulverised powders and passing them through #100 mesh sieve

75 2) Kneading: The Drug and HP- $\beta$  Cyclodextrin were triturated in different ratios using sufficient  
76 amount of water to form a thick paste, which was further kneaded for 15 minutes. Then it was allowed  
77 to dry in a hot air oven. The dried mass was then triturated using mortar and pestle, sieved through  
78 #100 mesh sieve, stored in a desiccator.

79 3) Solvent evaporation: To a drug solution in 70% v/v of Ethanol (10 ml) appropriate amount of HP- $\beta$   
80 Cyclodextrin, in different ratios were added. This solution was allowed to stir continuously using hot  
81 plate magnetic stirrer until the solvent evaporated. Solid dispersion thus obtained was then pulverized,  
82 sieved through mesh 100, stored in a desiccator.

### 83 **Characterization of Drug, Excipients and Solid Dispersion**

#### 84 **Fourier transform infra-red spectroscopy (IR)**

85 To study and confirm any drug-excipients interaction, Fourier Transform Infrared (FTIR)  
86 spectroscopy was performed. Initial spectra of the drug (OLM) and HP-  $\beta$ CD as control along with the  
87 solid dispersions were recorded. About 5-10 mg of test samples was used to study FTIR. FTIR  
88 spectroscopy of the test samples were performed by using the KBr scanning over wave number range  
89 of 4000–400  $\text{cm}^{-1}$  with FTIR spectrophotometer (FTIR-8001, Shimadzu, Japan) operated with IR  
90 Solution Software.

91 Drug (OLM), HP-  $\beta$ CD, physical mixture and Solid Dispersion were filled in amber colored vials  
92 sealed with bromo butyl rubber stoppers and kept in the environmental stability chamber (Remi Lab,  
93 Mumbai, India) for accelerated stability condition at  $40 \pm 2^\circ\text{C}$  temperature and  $75 \pm 5\%$  relative  
94 humidity for a period of 30 days. FTIR spectra of samples were obtained and spectra of solid  
95 dispersions were compared with the initial spectra of drug and excipients.

#### 96 **Differential Scanning Calorimetry (DSC)**

97 DSC curves of OLM, HP- $\beta$  Cyclodextrin, physical mixture and solid dispersion prepared by kneading  
98 method and solvent evaporation were obtained by using DSC (SHIMADZU DSC 60 PLUS, Japan) at  
99 a heating rate of  $5^\circ\text{C}/\text{min}$  from  $30^\circ\text{C}$  to  $300^\circ\text{C}$  in a nitrogen atmosphere. DSC curves of samples were

100 obtained, and spectra of solid dispersions compared with the initial spectra of drug and excipients to  
101 confirm the formation of inclusion complex.

### 102 **X-Ray Diffractometry (XRD)**

103 XRD was carried out at room temperature using a D/max 2500VL/PC powder X-ray diffractometer  
104 (Rigaku, Japan) was operated at 40 kV and 35 mA to analyse the physical nature of the drug and  
105 inclusion complexes. Test samples were scanned over a  $2\theta$  range of 3–40 with a step size of  $0.020^\circ$   
106 and a step time of 59.7 s.

### 107 **Scanning electron microscopy**

108 The SEM images of OLM, HP- $\beta$  Cyclodextrin and inclusion complexes prepared was observed by  
109 scanning electron microscope (JSM- 5510 (Jeol Ltd. Tokyo, Japan) with the accelerating voltage 10-  
110 kV and current of 25 mA. In this technique, the test samples were placed in the metal stub having two  
111 way adhesive tape and under reduced pressure of about 2.54 pa; it was coated with the very fine  
112 platinum film. The SEM images obtained were observed and analysed at different scale or resolutions.

### 113 **Determination of stability constant of the complex**

#### 114 **Phase solubility studies**

115 Higuchi and Connors method is a classical method for the Solubility studies, thus used here to  
116 perform phase solubility studies. In this method, excess drug was added to 30 ml of distilled water  
117 containing different molar concentrations of HP- $\beta$  cyclodextrin (0.002-0.01 M). These mixtures were  
118 shaken on the orbital shakers at 250 revolution per minute for 24 hrs at the room temperature. The  
119 samples were then removed from shaker and allowed to achieve equilibrium. The aliquots were then  
120 filtered through wattman filter paper. The samples were suitably diluted and analysed using UV  
121 spectrophotometer by measuring the absorbance at 254 nm wavelength.

#### 122 **Spectrophotometric studies**

123 Complex formation between OLM and HP- $\beta$  cyclodextrin was studied by keeping the OLM  
124 concentration fixed as  $2.445 \times 10^{-8}$  M whereas the concentration of HP- $\beta$  cyclodextrin was increased  
125 from 0.002 to 0.01M. The spectra of the drug were recorded on a UV-Vis spectrophotometer,  
126 Shimadzu (UV-1800 240V, Japan).The change in the absorbance of drug on the addition of various  
127 concentration of the HP- $\beta$  cyclodextrin were measured at 254 nm to evaluate the stability constant of  
128 the complex. A stability constant is a measure of the strength of the interaction which represent the  
129 equilibrium constant for the formation of a complex in solution.

130 Similarly, the UV spectra of the OLM, physical mixtures and inclusion complexes prepared by  
131 solvent evaporation and kneading method in different ratios were recorded and overlaid. The  
132 absorbance values of each sample containing fixed concentration of drug were measured at 254 nm to  
133 evaluate the enhanced solubility.

#### 134 **Dissolution and Accelerated Stability studies**

##### 135 **In vitro dissolution studies**

136 *In vitro dissolution studies* of pure drug, inclusion complexes and innovator (marketed product) were  
137 performed using LABINDIA DS 8000 USP paddle method at 50 rpm in 900 ml of 0.1 N HCl acid  
138 (pH 1.2) as dissolution media. The temperature was maintained at  $37 \pm 0.5^\circ\text{C}$ . 5 ml sample was  
139 withdrawn at the time intervals of 5, 10, 15, 30, 40 and 50 minutes, filtered through wattmann filter  
140 paper and analyzed spectrophotometrically at 254 nm. After each withdrawal of the sample an equal  
141 volume of prewarmed fresh medium at the same condition, was replaced into the dissolution media to  
142 maintain the constant volume throughout the test 3.

##### 143 **Accelerated stability testing**

144 Inclusion complexes were kept in sealed amber coloured bottles, protected from light and stored at  
145  $40^\circ\text{C}/75\%$  RH for 6 months. The products were analyzed for FTIR spectroscopy, drug content and  
146 dissolution after 6 month.

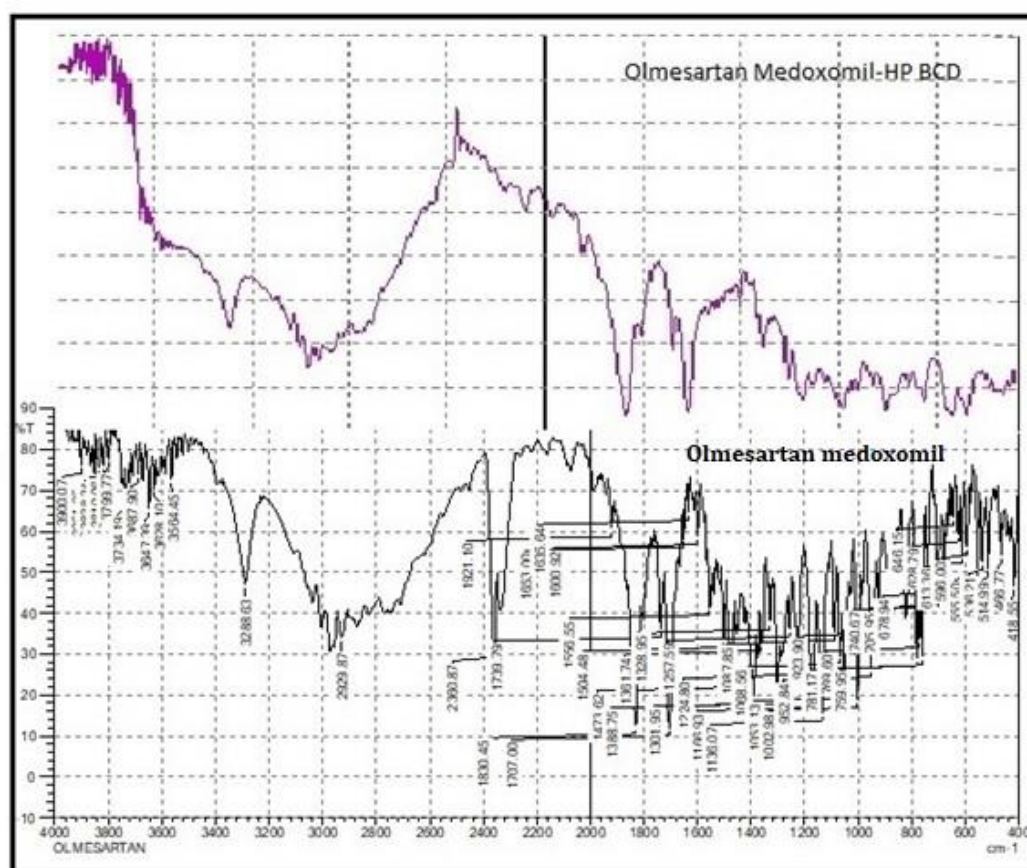
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148 **RESULTS AND DISCUSSION**

149 **Physical characterization**

150 **FT-IR Spectroscopy:**

151 The FT-IR spectra of pure Olmesartan medoxomil shows characteristic peaks at 2929.87, (C-H, str),  
152 1707, 1739.79, 1830.45 (C-O, str) and 3288.63 cm<sup>-1</sup> (N-H, str) The spectra of molecular inclusion  
153 complex of Olmesartan medoxomil with HP- $\beta$  cyclodextrin showed (Fig. 1) The characteristic peaks  
154 of Olmesartan medoxomil at initial conditions was maintained indicating that there is no chemical  
155 interaction between drug and excipients. Also, it was observed that there were no changes in the  
156 physical appearance of the samples.

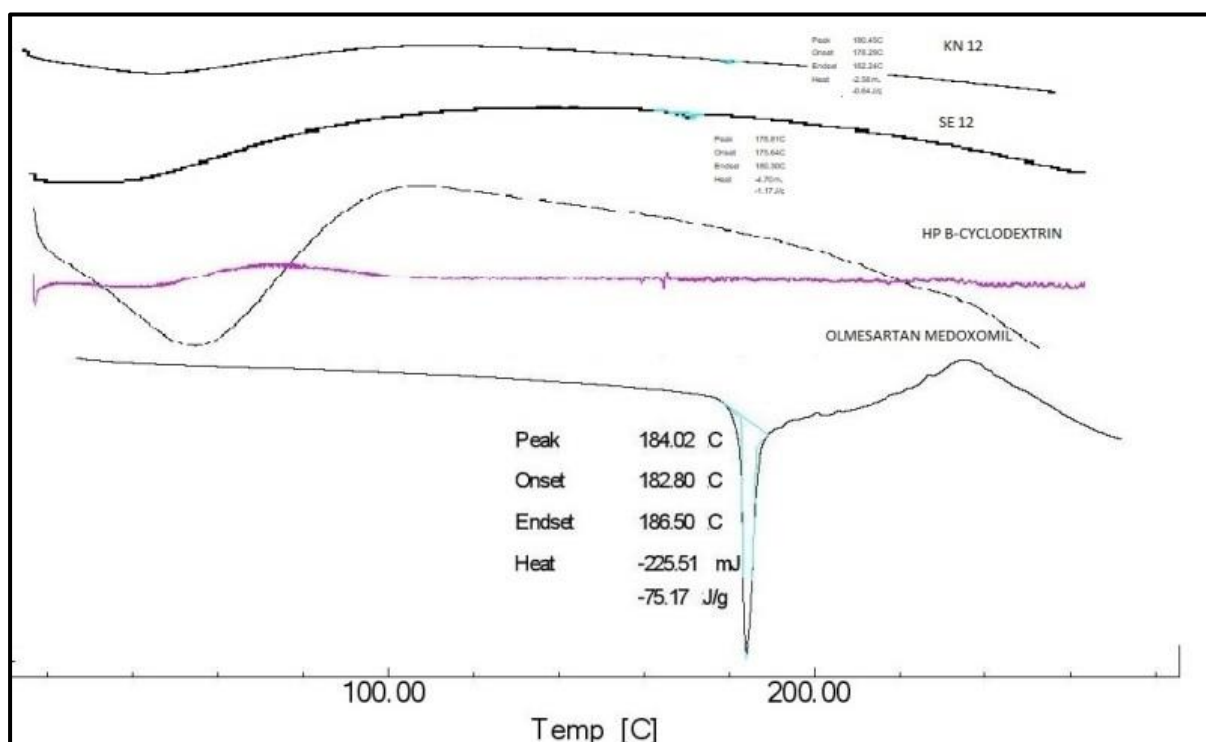


157  
158 **Figure 1.** FTIR spectra of Pure Olmesartan Medoxomil (OLM) and complex of Hydroxypropyl  $\beta$   
159 cyclodextrin (HP  $\beta$ CD) and Olmesartan Medoxomil (OLM).

160

161 **DSC studies**

162 The DSC thermograms of Olmesartan medoxomil, HP- $\beta$  cyclodextrin and the inclusion complexes are  
163 shown in Fig. 2. The DSC curve of HP- $\beta$  cyclodextrin shows a broad endothermic peak which was  
164 due to its amorphous nature. The sharp peak of Olmesartan medoxomil appeared at 184.02 ° C,  
165 whereas no such peak was observed in inclusion complex prepared with HP- $\beta$  cyclodextrin suggesting  
166 that Olmesartan was molecularly dispersed. It might have been no longer available in a crystalline  
167 state, and was converted into the amorphous state.



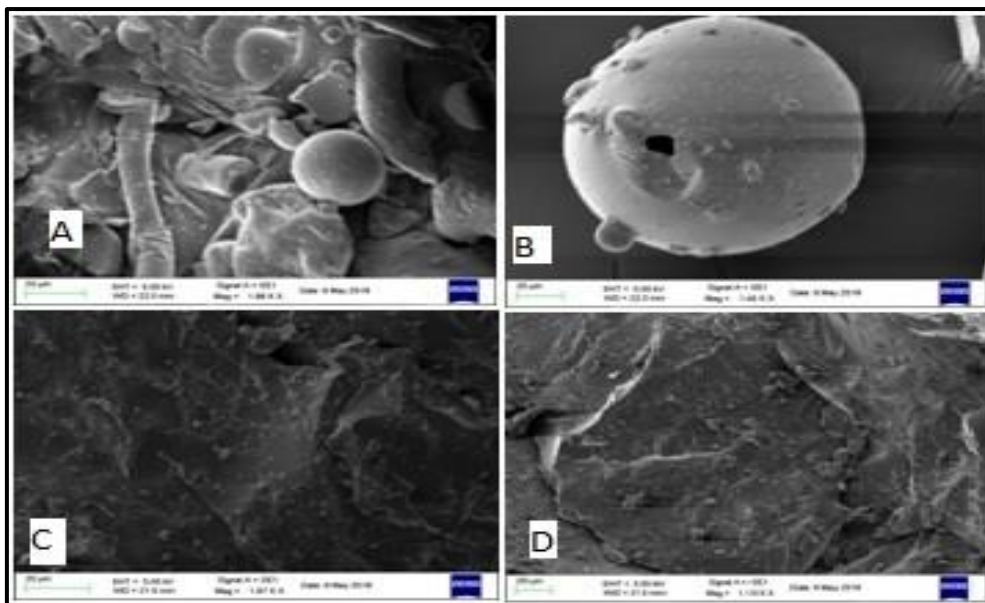
168  
169 **Figure 2.** DSC curves of Pure Olmesartan Medoxomil (OLM), Hydroxypropyl  $\beta$  cyclodextrin (HP  
170  $\beta$ CD), Olmesartan Medoxomil (OLM): Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ CD) kneaded complex  
171 (1:2), and Olmesartan Medoxomil (OLM): Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ CD) solvent  
172 evaporated complex (1:2).

173 **SEM ANALYSIS**

174 The surface morphological features of OLM, HP-  $\beta$ CD, and inclusion complexes are shown in figure  
175 3. The complexes appeared as agglomerates rather than individual crystalline structures. This change



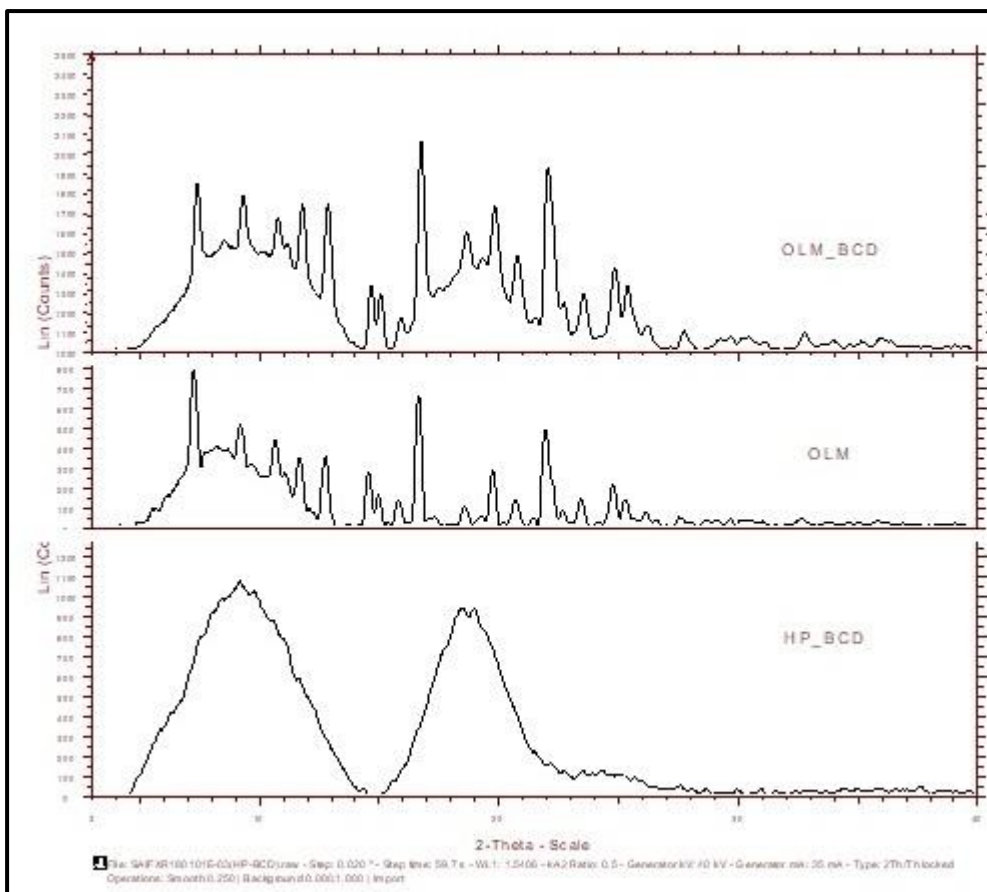
176 of the particle shape was indicative of a conversion of solid crystalline state to amorphous state of the  
177 complexes.



178  
179 **Figure 3.** Scanning electron microscopic images of A- OLM and HP-  $\beta$  CD, B- HP-  $\beta$  CD,  
180 C -Inclusion complex of HP  $\beta$ -CD (SE 12) and D- Inclusion complex of HP  $\beta$ -CD (KN 12)

### 181 XRD analysis

182 The crystallinity of the pure drug and inclusion complexes could be studied through XRD patterns as  
183 shown in the fig 4. It was observed that pure OLM exhibited a characteristic peaks in the  $2\theta$  angle  
184 located at  $7^\circ$ ,  $9^\circ$ ,  $11^\circ$ ,  $12^\circ$ ,  $14^\circ$ ,  $16^\circ$ ,  $19^\circ$ , and  $21^\circ$  which indicates the crystalline nature of OLM.  
185 Whereas, X-ray diffractometer graph of HP- $\beta$ CD shows no characteristic peak, which indicates the  
186 amorphous nature of the carrier. When the inclusion complex prepared by solvent evaporation and  
187 kneading technique were analysed, they also showed completely diffused diffraction patterns  
188 indicating conversion of crystalline form into amorphous. This suggests that OLM was entrapped  
189 within the hydrophobic cavity of HP $\beta$ CD. Absence of crystallinity and developed amorphous nature  
190 of the evaporated and kneaded complexes of OLM: HP $\beta$ CD suggests the improved solubility and  
191 dissolution profile of OLM in gastrointestinal media.

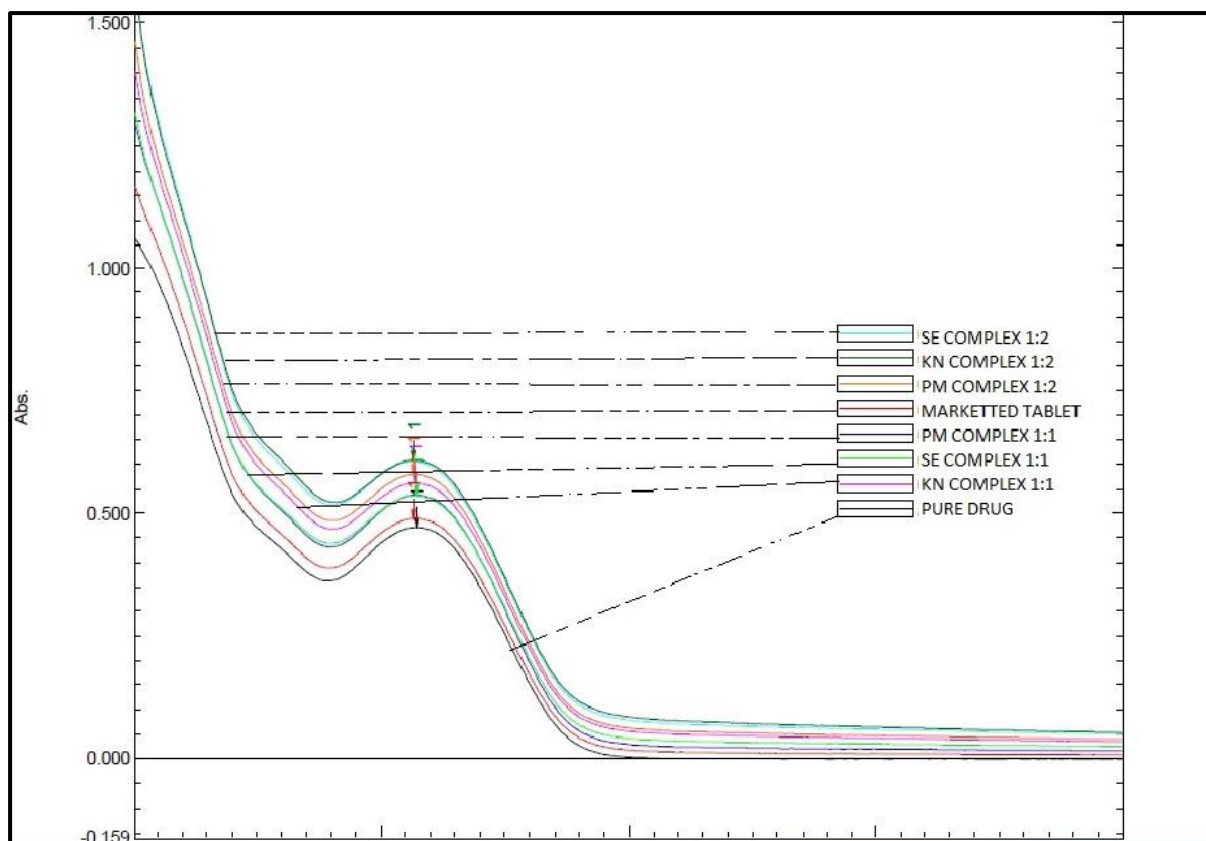


192

193 **Figure 4.** X-ray diffraction patterns of Pure Olmesartan Medoxomil (OLM), Hydroxypropyl  $\beta$   
 194 cyclodextrin (HP  $\beta$ CD) and kneading complex of Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ CD) and  
 195 Olmesartan Medoxomil (OLM)

196 **UV Spectroscopy**

197 The Olmesartan medoxomil solution in the presence of increasing molar concentration of HP- $\beta$ CD  
 198 was scanned and analysed on UV spectrophotometer. The spectra of each solution recorded are shown  
 199 in Fig. 5. There are changes observed in the peak intensity in spite of addition of fixed amount of drug  
 200 in solution. This may be the result of transfer of the drug molecule from water to the HP- $\beta$ CD cavity,  
 201 resulting in enhanced solubility of drug as indicated by the increased peak intensity 5.



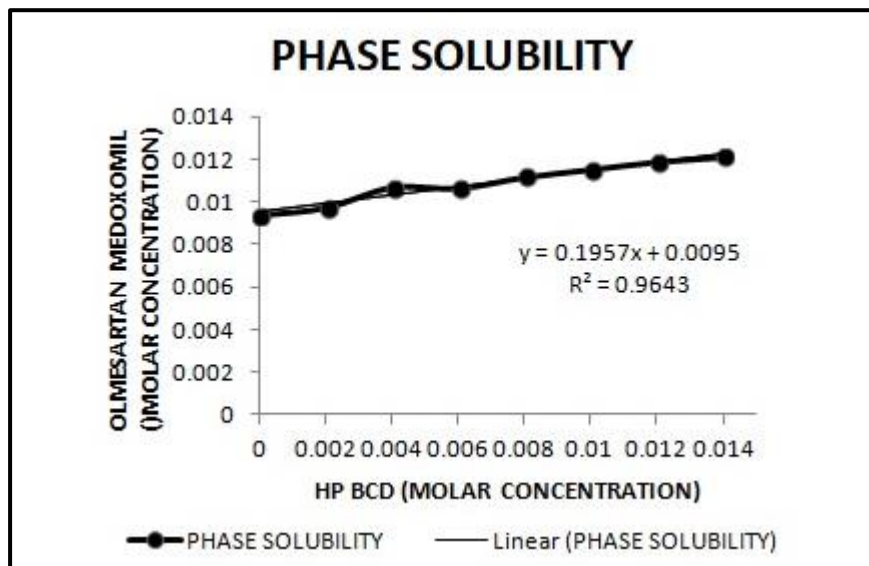
202  
 203 **Figure 5.** Overlay Spectra of Olmesartan Medoxomil (OLM) and various complexes prepared with  
 204 Hydroxypropyl-β cyclodextrin (HP βCD).

205 **Determination of Stability Constants**

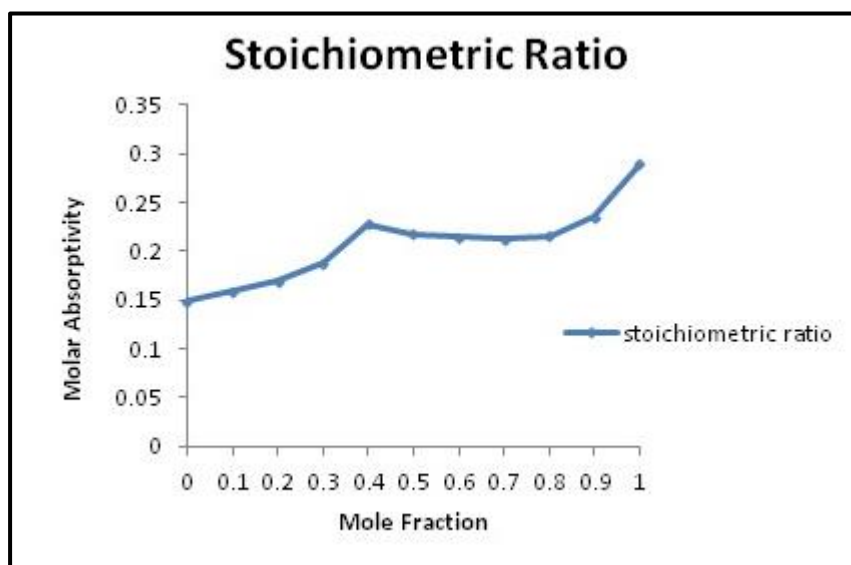
206 The complexing behaviour of Olmesartan Medoxomil with the HP-β cyclodextrin was studied by the  
 207 phase-solubility method. The phase solubility diagram for the inclusion complex formation between  
 208 Olmesartan medoxomil and HP-β cyclodextrin is shown in Fig. 6. It was observed from the phase  
 209 solubility diagram that the aqueous solubility of Olmesartan medoxomil have increased linearly, as a  
 210 function of HP-β cyclodextrin concentration. The graph shows the value with a slope 0.1957 (r=  
 211 0.9643). The stability constant ( $K_c$ ) of Olmesartan medoxomil was calculated as  $11858.15 \text{ M}^{-1}$  from  
 212 the plot of the phase solubility diagram and using the equation

213 
$$K_c = \frac{\text{Slope}}{S_0(1-\text{Slope})} \quad (1)$$

214 where, So is the solubility of drug. It can be concluded from the results obtained by using method of  
215 continuous variation (Job's Plot) as shown in fig.7 that the increase in solubility observed was due to  
216 the formation of a 1:2 M inclusion complex.



217  
218 **Figure 6.** Phase solubility diagram of Olmesartan Medoxomil (OLM) in Hydroxypropyl  $\beta$   
219 cyclodextrin (HP  $\beta$ CD) solution.



220  
221 **Figure 7.** Job's Plot showing stoichiometric ratio of Olmesartan Medoxomil (OLM) in  
222 Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ CD) molecular inclusion complex.

223 Absorption spectra used to confirm the formation of inclusion complex and to compare the degree of  
224 enhancement of solubility. In this study, absorption spectra of HP- $\beta$  cyclodextrin, Olmesartan  
225 medoxomil, physical mixture and inclusion complex were taken into consideration. In the absorption  
226 spectra for Olmesartan medoxomil and the physical mixture there is only a slight increase in the  
227 absorbance peak along the wavelength was recorded which may be due to the higher wetting effect of  
228 the available HP- $\beta$  cyclodextrin with drug. Whereas, inclusion complex showed much more rise in  
229 intensity at all points of wavelength, this may be due to the inclusion or molecular dispersion of drug  
230 in HP- $\beta$  cyclodextrin cavity.

231 Stability constant was determined using the Benesi–Hildebrand equation

232

$$233 \quad \frac{1}{\Delta A} = \frac{1}{(D)} Kc \Delta \epsilon X \frac{1}{(CD)} + \frac{1}{(D)} \Delta \epsilon \quad (2)$$

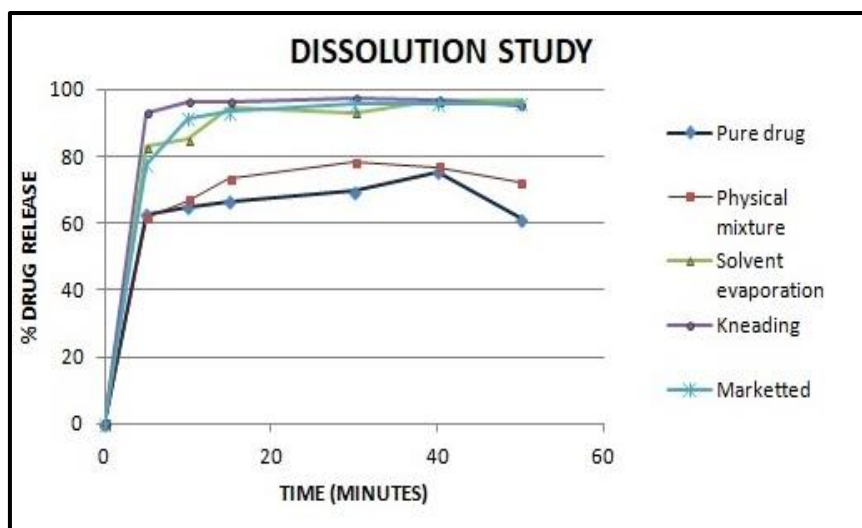
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235 where  $\Delta A$  is the difference of absorbance at 254 nm, (CD) the HP-  $\beta$  cyclodextrin concentration, (D)  
236 the drug concentration (constant), and  $\Delta \epsilon$  is the difference in the molar absorptivity values between  
237 the complex and the free drug. The Kc value was calculated to be 5531.12 M<sup>-1</sup>. It was found that the  
238 stability constant which was calculated by a spectral shift technique was relatively small compared to  
239 that obtained by the phase solubility studies. However, from the literature chaudhary et.al it was noted  
240 that the use of the spectral shift technique is not the method of choice for a reliable determination the  
241 drug with low aqueous solubility, because the difference in the absorptivity values will be too small 2.  
242 In contrast, the other phase solubility method provides a much better and reliable results in these  
243 cases.

#### 244 **In vitro dissolution studies**

245 The dissolution curves of Pure drug, Physical mixtures, Solvent evaporation and Kneading solid  
246 dispersions in 0.1 N HCl (pH=1.2) at 37  $\pm$  0.5°C are shown in Fig. 8 and the corresponding values are  
247 given in Table 2. From the results obtained, it was observed that all the binary systems of Olmesartan  
248 Medoxomil demonstrated higher dissolution rates than pure drug and its corresponding physical  
249 mixtures. The physical mixtures also showed improved dissolution rate as compared with OLM but

250 the highest drug release is observed for molecular inclusion complexes prepared by Kneading method.  
 251 It is evident that the inclusion complex improved the dissolution rate of OLM to the greatest extent.  
 252 Amongst all preparations, OLM: HP- $\beta$ CD 1:2 inclusion complex produced by kneading method  
 253 showed highest dissolution rate of about 96.73 % of drug in pH 1.2 at the end of 30 min.



254  
 255 **Figure 8.** Dissolution profile of Olmesartan Medoxomil, prepared Olmesartan Medoxomil (OLM) :  
 256 Hydroxypropyl  $\beta$  cyclodextrin (HP  $\beta$ CD) molecular inclusion complexes and Marketed preparation in  
 257 0.1 N hydrochloric acid.

258 **Table 1.** Olmesartan medoxomil content in HP $\beta$ CD inclusion complex systems prepared.

Method	Formulation code	Complex system	Olmesartan medoxomil content (%)
Physical mixing	PM 11	Olmesartan medoxomil: HP- $\beta$ CD (1:1)	98.52
	PM 12	Olmesartan medoxomil: HP- $\beta$ CD (1:2)	98.64
	PM 13	Olmesartan medoxomil: HP- $\beta$ CD (1:3)	98.45
Kneading	KN 11	Olmesartan medoxomil: HP- $\beta$ CD (1:1)	99.21
	KN 12	Olmesartan medoxomil: HP- $\beta$ CD (1:2)	99.10
	KN 13	Olmesartan medoxomil: HP- $\beta$ CD (1:3)	98.78
Solvent evaporation	SE 11	Olmesartan medoxomil: HP- $\beta$ CD (1:1)	98.11
	SE 12	Olmesartan medoxomil: HP- $\beta$ CD (1:2)	98.24
	SE 13	Olmesartan medoxomil: HP- $\beta$ CD (1:3)	98.37

PM 11 ; Physical Mixture (1:1), PM 12 ; Physical Mixture (1:2), PM 13 ; Physical Mixture (1:3), KN 12; Kneading Complex (1:1),KN 12; Kneading Complex (1:2), KN 13; Kneading Complex (1:3), SE 11; Solvent Evaporation Complex (1:1) ,SE 12; Solvent Evaporation Complex (1:2), SE 13; Solvent Evaporation Complex (1:3).

259

260 **Table 2.** Dissolution profile of Olmesartan medoxomil in 0.1 N hydrochloric acid

TIME (min)	Mean percentage of drug dissolved ( $\pm$ SD)* n=3				
	PD	PM 12	SE12	KN 12	INN
5	62.52(1.56)	61.69(1.01)	82.86(1.19)	92.88(0.59)	77.56(1.14)
10	64.66(1.75)	66.88(1.14)	85.25(1.63)	96.46(1.53)	91.47(1.01)
15	66.27(1.68)	73.32(1.11)	94.73(0.73)	96.29(1.01)	93.32(1.01)
30	69.52(0.83)	78.08(1.20)	93.08(1.40)	97.40(1.17)	95.54(1.5)
40	75.09(1.63)	76.86(1.23)	96.61(1.15)	96.73(0.97)	95.79(0.74)
50	61.18(1.00)	71.92(2.14)	96.51(1.00)	95.09(1.57)	95.84(1.37)
DE <sub>15</sub> %	59.66	61.75	77.23	83.52	77.79
Similarity factor	36.94	42.01	78.37	65.59	Reference
PD, Pure Drug ; PM 12 ; Physical Mixture (1:2), SE 12; Solvent Evaporation Complex (1:2), KN 12; Kneading Complex (1:2), INN, Marketed Tablet (Powdered)					

261

262 Dissolution efficiency (% DE) is the area under the dissolution curve between time point's  $t_1$  and  $t_2$   
 263 expressed as a percentage of the curve at maximum dissolution,  $y_{100}$ , over the same time period and  
 264 is expressed by the following expression:

265 **Dissolution efficiency (DE)** = 
$$\frac{\int_{t_1}^{t_2} y dt}{y_{100}(t_2 - t_1)} \quad (3)$$

266 DE<sub>15</sub>% values of PD, PM12, SE12, KN12 and INN are found to be 59.66, 61.75, 77.23, 83.52  
 267 and 77.79 respectively.

268 **Accelerated stability testing:**

269 Accelerated stability testing results showed no significant variations neither in the content of drug nor  
 270 in dissolution profiles after storage at 40 °C/75% RH for 6 months. Therefore, all these results imply

271 that the prepared molecular inclusions are stable over the storage period and no effect on its stability  
272 on storage.

273 In the present study it was clearly observed that OLM immediate release inclusion complex can be  
274 effectively produced by processing via HP- $\beta$ CD with enhanced solubility and Dissolution Rate. The  
275 results of the studies also indicated the formation of HP  $\beta$ CD- Olmesartan medoxomil molecular  
276 inclusion complex prepared by kneading method at different ratios of drug and HP- $\beta$ CD. The  
277 inclusion complex had enhanced dissolution rate of OLM to a greater extent compared to the  
278 corresponding solvent evaporation, physical mixture and pure OLM,. Further, it was noted that OLM-  
279 incorporated molecular inclusions in 1:2 ratio aqueous solution provides the highest dissolution and  
280 solubility values compared to the marketed product and OLM pure drug, which indicate that the oral  
281 bioavailability of OLM might be improved. Kneading method was the most effective method in terms  
282 of OLM solubility and dissolution. This enhanced dissolution from the inclusion complexes may be  
283 the result of increased hydrophilicity and wetting effect that results from the interaction between the  
284 drug and the carrier. The another reason for the rapid dissolution of OLM from inclusion complexes  
285 may be attributed to the entrapment of drug in the hydrophilic carrier matrix of carrier HP- $\beta$ CD and a  
286 decrease in the crystallinity of drug as observed in XRD and DSC studies. Further, in the mechanism  
287 of quick dissolution of drug from the inclusion complexes could be due to the soluble carrier  
288 dissolving into the dissolution medium and allowing the entrapped drug release to the medium in the  
289 form of very fine particles.

290 Thus, we can conclude that HP- $\beta$ CD is useful as a primary carrier in the formulation of molecular  
291 inclusion complex of the drug (OLM) prepared by kneading method so as to enhance its solubility and  
292 dissolution rate.

### 293 **Acknowledgement**

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295 drug and Rouquette, France for providing gift sample of KLEPTOSE<sup>®</sup> HP- $\beta$ CD for this research.

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297 providing a fellowship.

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